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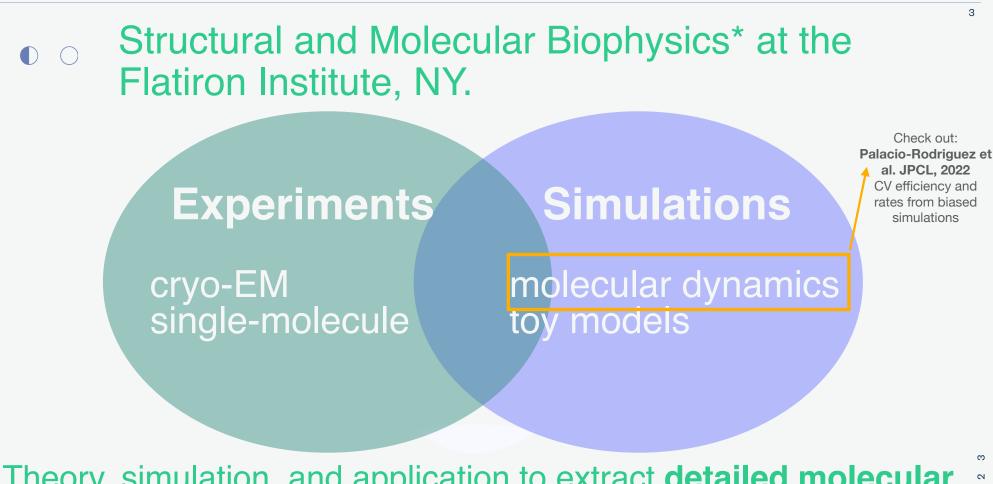
Ensemble reweighting using cryo-EM particles

Pilar Cossio Center for Computational Mathematics Flatiron Institute, NY

Outline:

- 1. Motivation
- 2. Cryo-EM as a single-molecule technique
- 3. Free-energies profiles along a path
- 4. Generalization: Ensemble Reweighting
- 5. Conclusions and perspectives: biasing MD...

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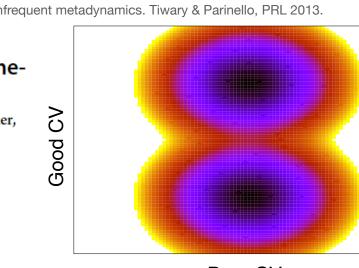
Theory, simulation, and application to extract detailed molecular mechanisms: free energies and dynamics.

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* Co-leadership with Sonya Hanson





Poor CV

 $k(t) = k_{\rm pre} e^{-\beta \Delta G^{\ddagger} + \beta \gamma V_{\rm MB}(t)} = k_0 e^{\beta \gamma V_{\rm MB}(t)}$

Read Online

Scales the bias

Fitting parameters: γ in [0,1] & γ =1 -> good CV

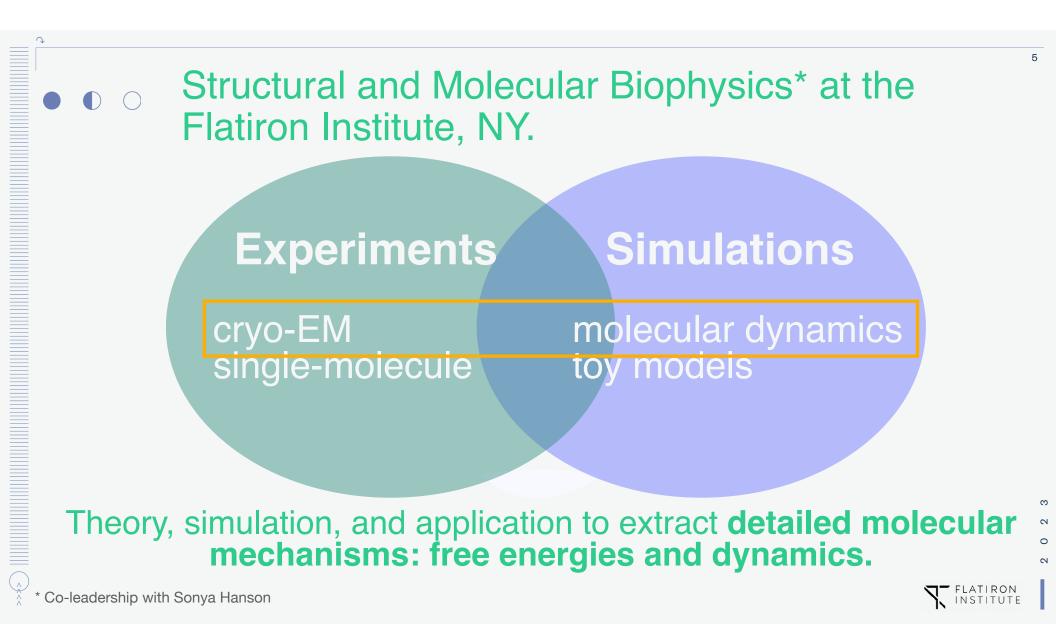
We can extract the unbiased rate (for any CV) and have a new measure of the efficiency of the CV.



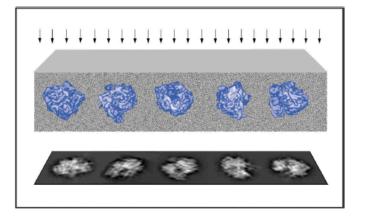
We introduce a measure of the *efficiency of CV* (y):

Karen Palacio-Rodriguez,[†] Hadrien Vroylandt,[†] Lukas S. Stelzl, Fabio Pietrucci, Gerhard Hummer, and Pilar Cossio*

Cite This: J. Phys. Chem. Lett. 2022, 13, 7490-7496

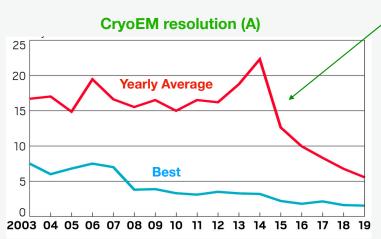


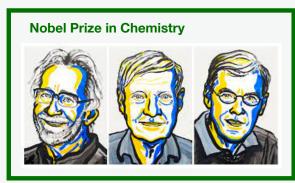
Cryo-EM images of biomolecules have two main unknowns: pose and conformation



Projection direction? -Conformation? 6

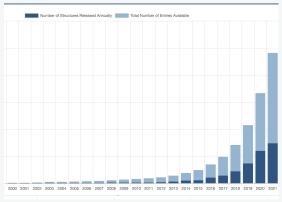
The CryoEM resolution revolution





Dubochet, Frank & Henderson

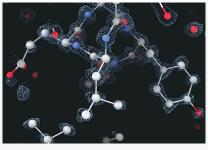
CryoEM structures in the PDB



NEWS AND VIEWS 21 October 2020

Cryo-electron microscopy reaches atomic resolution

2020 ~1.2A Resolution!



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However, CryoEM grounds have been shaken:



Where is the field going?

Faster, better and cheaper structure determination.



Conformational variability, free energies and environment

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CryoEM: conformational variability, free energies and environment

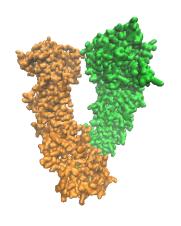
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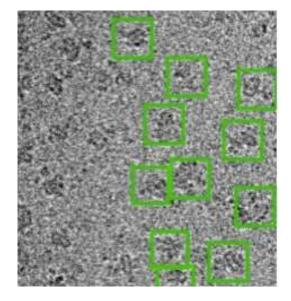
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Conformational heterogeneity from singleparticle cryo-EM

Freezing is done very rapidly*.

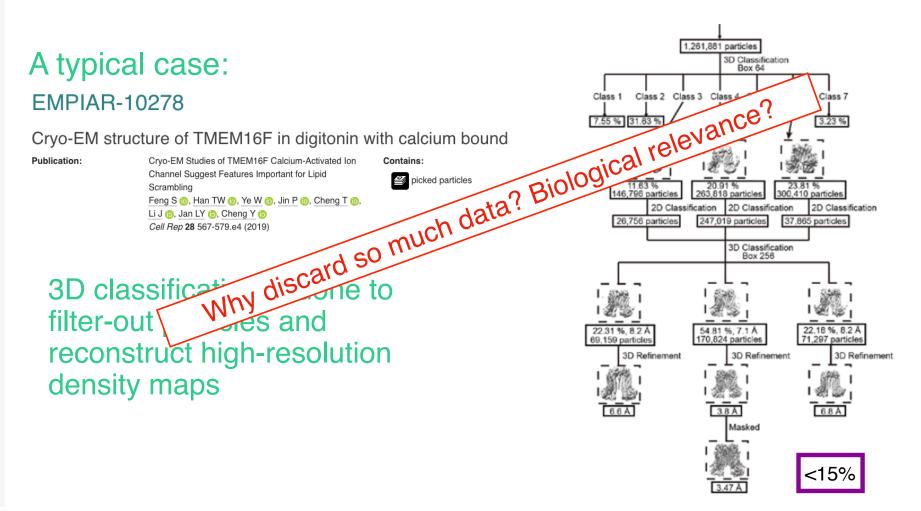
It's possible to trap individual conformations!!

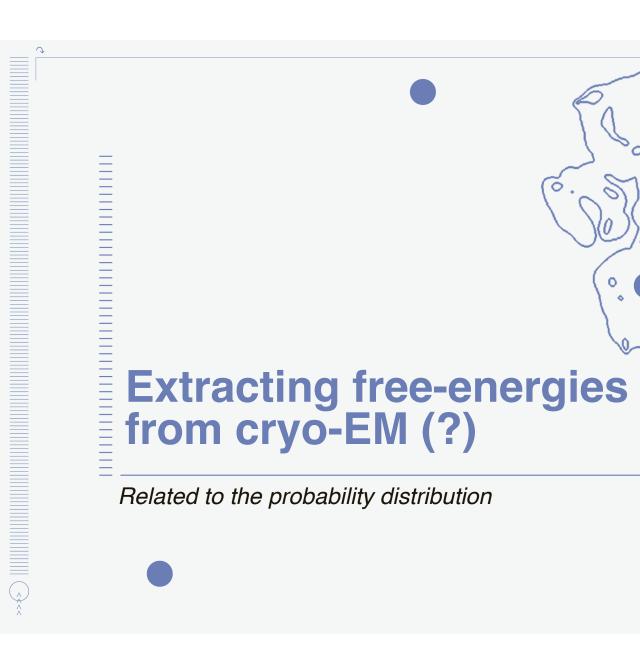




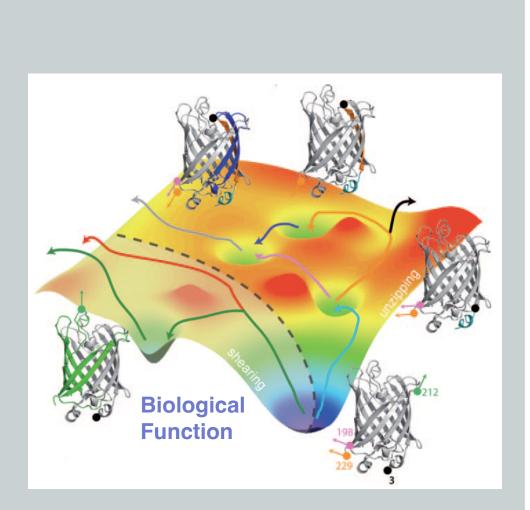
More information than just an average.

Dealing with cryoEM heterogeneity









Bertz et al. Angew. Chem. 2008, 47, 8192 - 8195

The free energy (FE) is related to the probability distribution of the configurations.

A configuration $x \in \mathbb{R}^{3N}$ and the Boltzmann factor gives the probability

$$\rho(x) = \frac{1}{Z} e^{-\beta H(x)} \sum_{\text{Hamiltonian}}$$

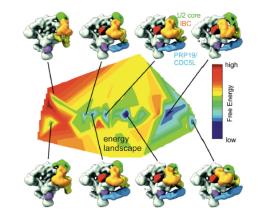
Too complex! Project along some collective variables (*s*):

$$\rho(s) = \int \delta(S(x) - s)\rho(x)dx = \frac{1}{Z_1} e^{-\beta G(s)}$$

Free energy

Free-energy surfaces from cryo-EM

Principal component analysis (PCA)



Machine learning & diffusion maps

Trajectories of the ribosome as a (2014)Brownian nanomachine

Ali Dashti^{a,1}, Peter Schwander^{a,1}, Robert Langlois^b, Russell Fung^a, Wen Li^b, Ahmad Hosseinizadeh^a, Hstau Y. Liao^b, Jesper Pallesen^{c,2}, Gyanesh Sharma^{b,3}, Vera A. Stupina^d, Anne E. Simon^d, Jonathan D. Dinman^d, Joachim Frank^{b,c,4}, and Abbas Ourmazd^{a,1,4}

⁽²⁰²⁰⁾ Retrieving functional pathways of biomolecules from single-particle snapshots

Ali Dashti^{1,9}, Ghoncheh Mashayekhi^{1,9}, Mrinal Shekhar (2,3, Danya Ben Hail⁴, Salah Salah^{4,5,6}, Peter Schwander ¹, Amedee des Georges ^{4,5,6^{III}}, Abhishek Singharoy ^{3^{III}}, Joachim Frank ^{7,8^{III}} & Abbas Ourmazd_☉ ^{1⊠}

Energy Landscape of the SARS-CoV-2 Reveals

(2021)

Extensive Conformational Heterogeneity

Ghoncheh Mashayekhi,^{1#} John Vant,^{2#} Abhishek Singharoy²*, Abbas Ourmazd¹*

Structure and Conformational Dynamics of the (2018)Human Spliceosomal B^{act} Complex Authors

David Haselbach, Ilya Komarov, Dmitry E. Agafonov, ..., Berthold Kastner, Reinhard Lührmann, Holger Stark

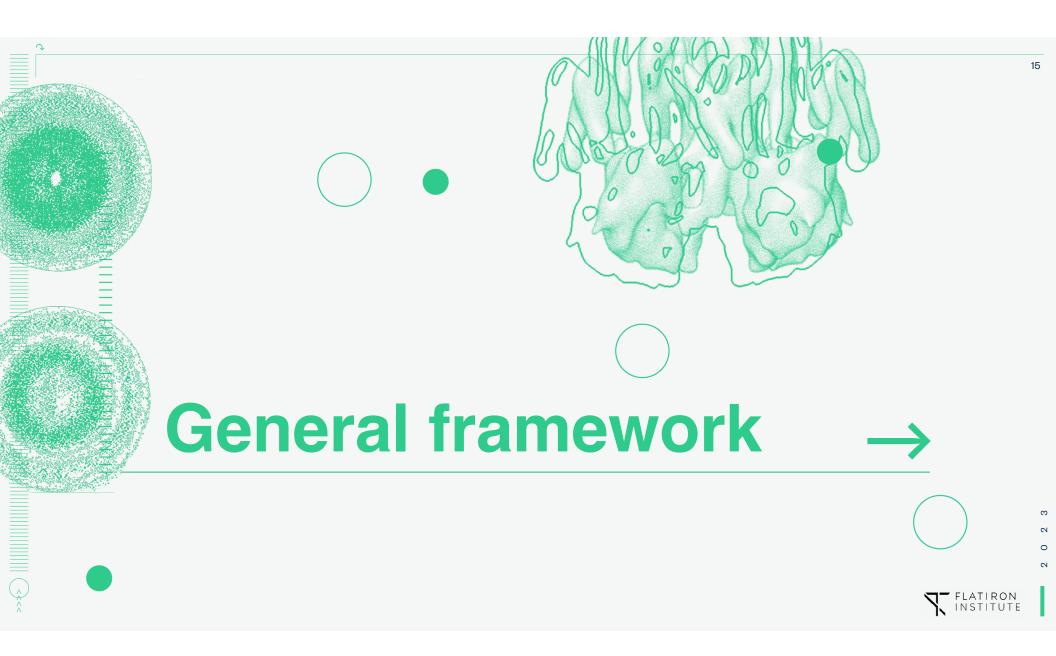
(2020)

Energy landscape of domain motion in glutamate dehydrogenase deduced from cryo-electron microscopy

Mao Oide^{1,2}, Takayuki Kato³, Tomotaka Oroguchi^{1,2} and Masayoshi Nakasako^{1,2}

Ribosome dynamics and tRNA movement by time-resolved electron cryomicroscopy

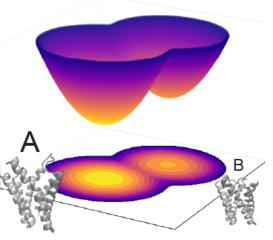
Niels Fischer¹, Andrey L. Konevega^{2,3}, Wolfgang Wintermeyer², Marina V. Rodnina² & Holger Stark¹



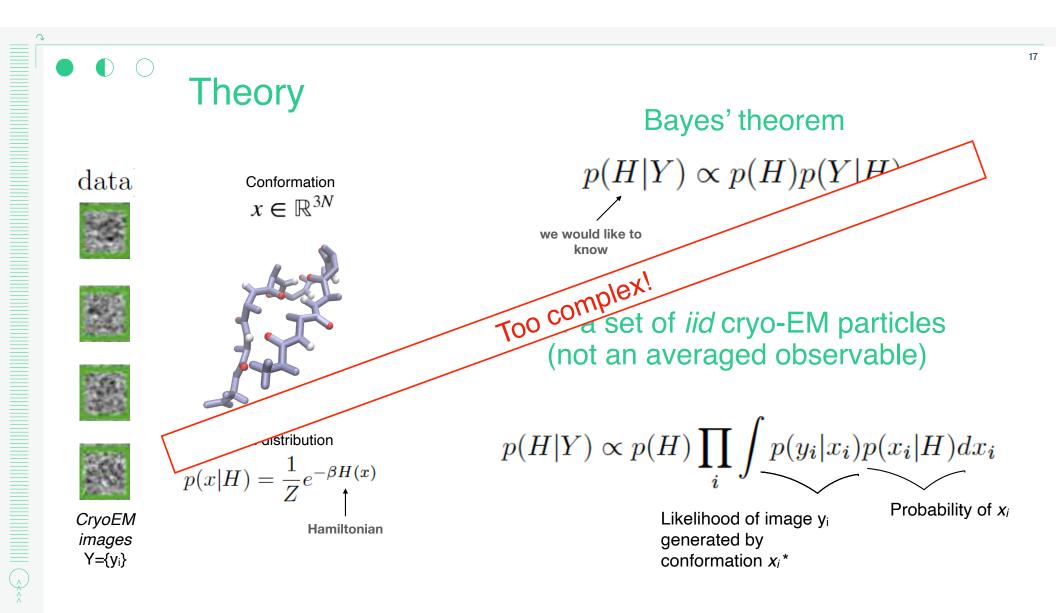
● ○ Free energies from CryoEM

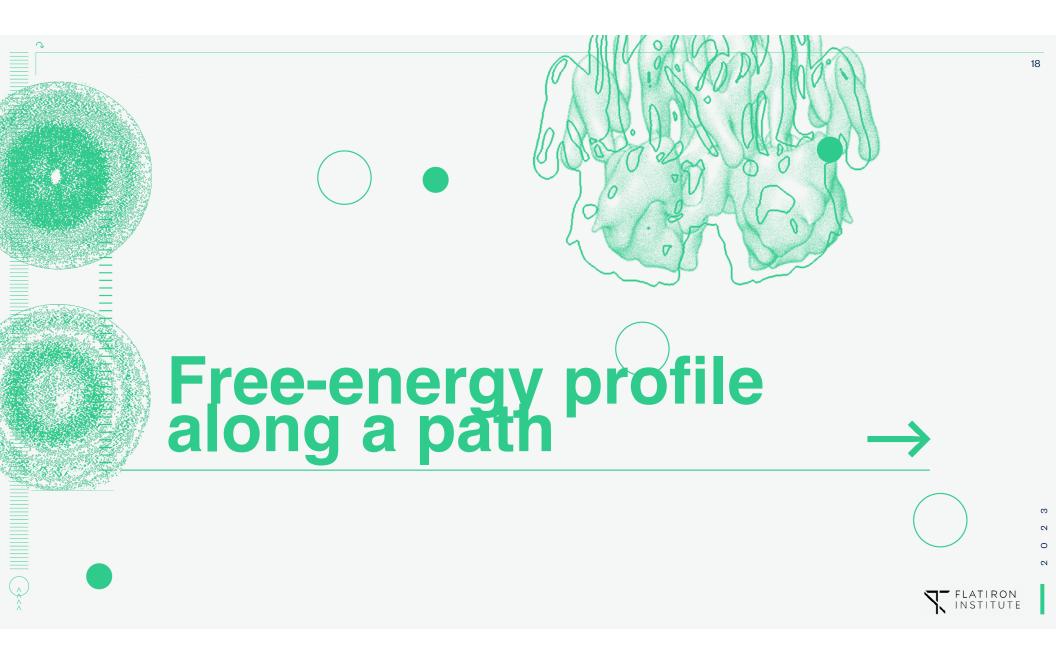
Inference of free energies given cryoEM particles.

Initial



Conformational space $x \in \mathbb{R}^{3\Lambda}$ Ensemble of models (e.g. AlphaFold, MD trajectory)





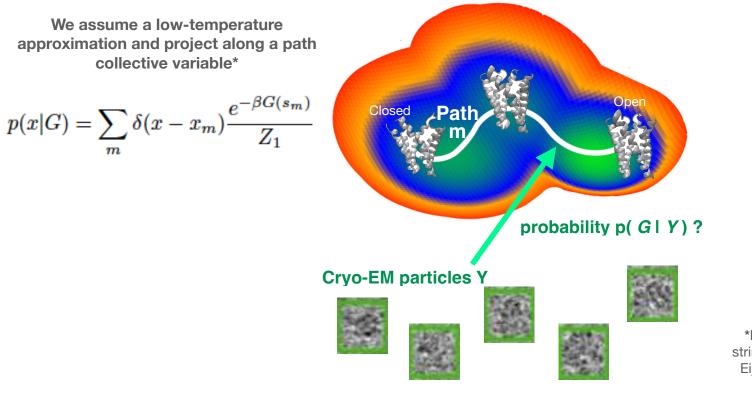
• • Cryo-BIFE

Cryo-EM Bayesian Inference of Free Energy

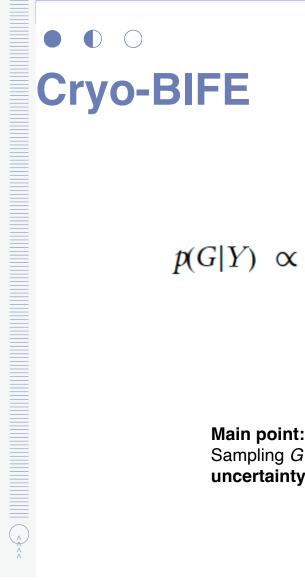
Article Open Access Published: 01 July 2021

A Bayesian approach to extracting free-energy profiles from cryo-electron microscopy experiments

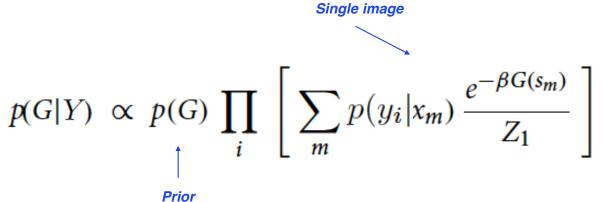
Julian Giraldo-Barreto, Sebastian Ortiz, Erik H. Thiede, Karen Palacio-Rodriguez, Bob Carpenter, Alex H. Barnett & Pilar Cossio 🖂



*Inspired by MD methods like the string method (Weinan, Ren, Vanden-Eijnden - Physical Review B, 2002)



For multiple images Y, the posterior reduces to



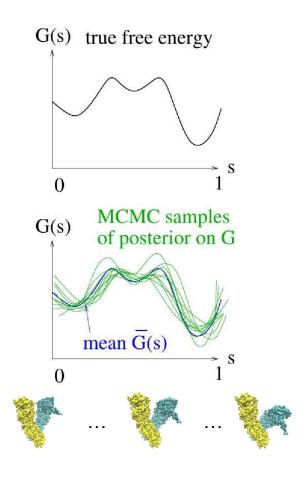
Probability of a *G* given the particles

Main point:

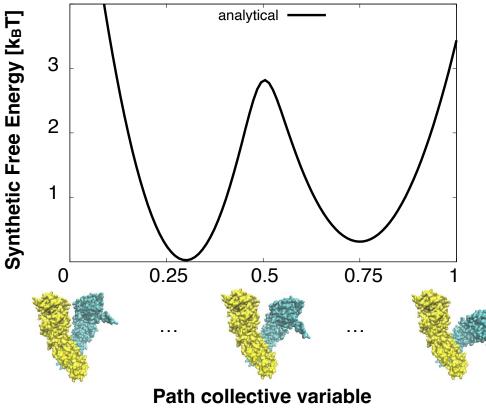
Sampling G from p(G | Y), allows us to calculate its mean and **uncertainty** at each node given the cryo-EM particles.

Key ideas:

- Particle reconstructions are **not** crucial.
- We assume that we have a sufficiently good transition path of 3D structures (by MD, Alphafold etc).
- We use MCMC sampling (e.g. in STAN) to extract the expected G and its uncertainty.



ID Hsp90 synthetic data

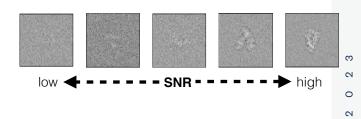


Path calculation:

- 20 nodes (along the single degree of freedom)
- 2 orientation-rounds of BioEM

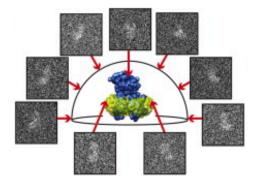
Synthetic images:

- ~13300 particles
- 128x128 pixels
- pixel size 2.2 A
- Coarse-grained residues With uniformly distributed:
- random orientations
- random defocus [0.5,3] micro-m
- random signal-to-noise ratio (SNR) [0.001, 0.1]

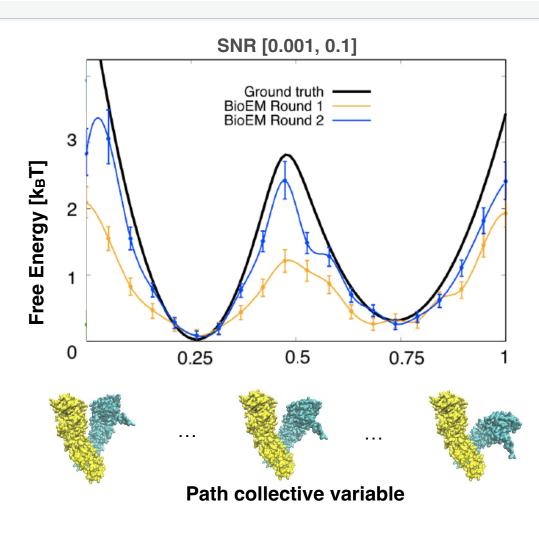


Projection direction accuracy is important

The projection direction is unknown.



Nogales & Scheres, Mol Cell 2015



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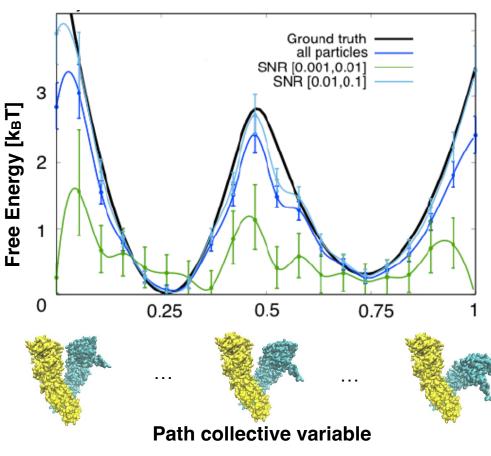
Orient. Round 2

● ● ○ Signal-to-noise ratio (SNR)

A poor FE profile is recovered from the low SNR group

High SNR provides a slightly better profile than using all particles.

Adding "bad" particles does not hinder the FE recovery (if there are some "good" ones in the group).



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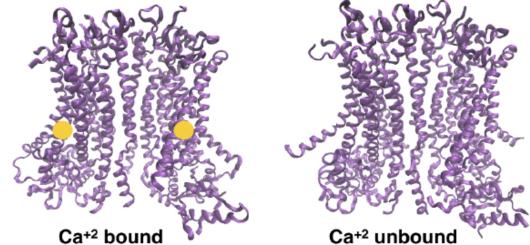
Real data:TMEM16F a calcium-activated ion channel

Cryo-EM Studies of TMEM16F (2019) Calcium-Activated Ion Channel Suggest Features Important for Lipid Scrambling

Shengjie Feng,^{1,4} Shangyu Dang,^{2,4} Tina Wei Han,¹ Wenlei Ye,¹ Peng Jin,¹ Tong Cheng,¹ Junrui Li,² Yuh Nung Jan,^{1,2,3} Lily Yeh Jan,^{1,2,3,*} and Yifan Cheng^{2,3,5,*}

¹Department of Physiology, University of California, San Francisco, San Francisco, CA 94158, USA ²Department of Biochemistry and Biophysics, University of California, San Francisco, San Francisco, CA 94158, USA ³Howard Hughes Medical Institute, University of California, San Francisco, San Francisco, CA 94158, USA

Generated from two different data sets: with and without Ca+2



All particles available: EMPIAR 10278

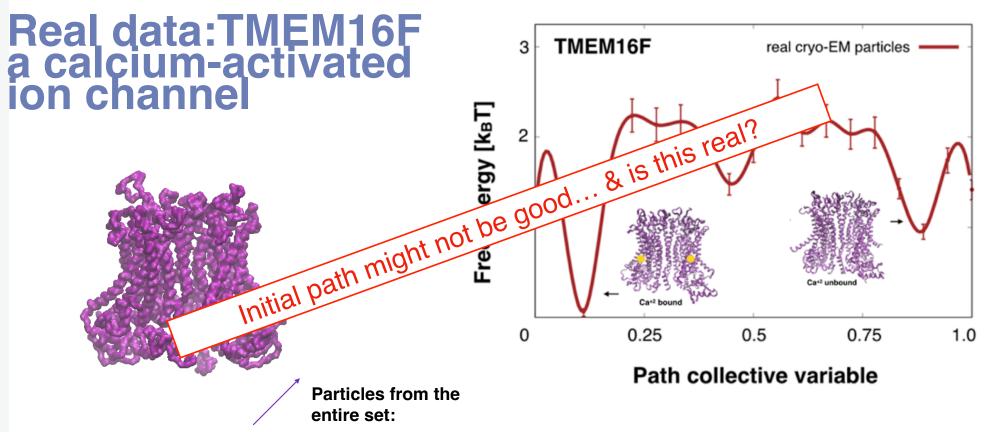
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Is there a population of the Ca+2-unbound state in the Ca+2-bound set?



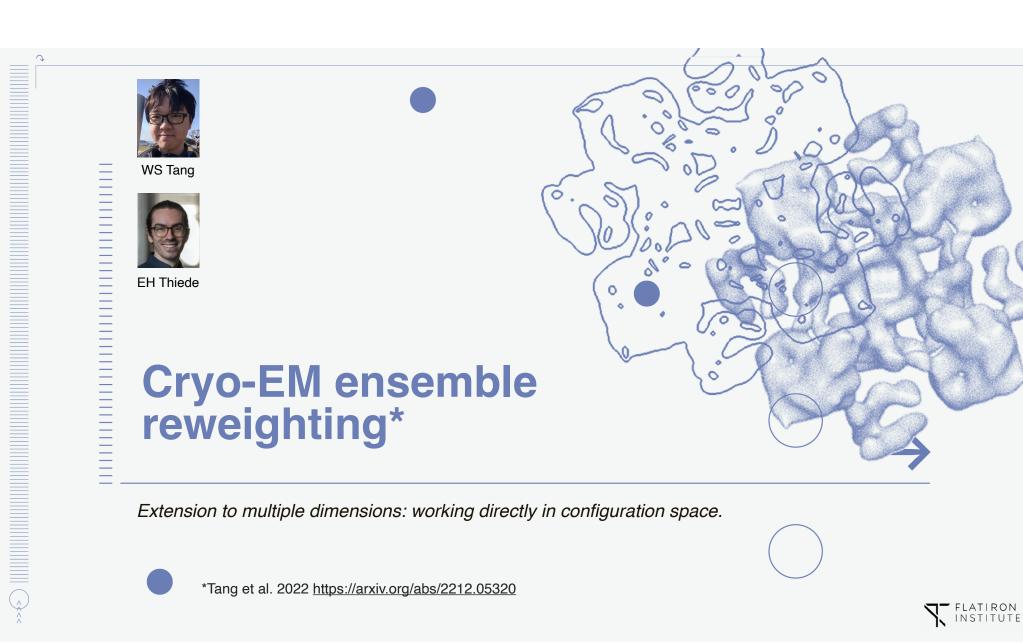


- EMPIAR 10278
- 15000 images
- 256x256 pixels
- pixel size 1 A

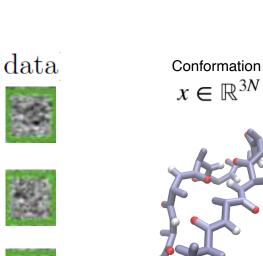
Opening Pandora's box

- Extension to many dimensions
- Experimental validation system
- How to optimize the molecular path?
- Comparing many models vs many images optimization.

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CryoEM images $Y = \{y_i\}$

Theory



Boltzmann distribution

 $p(x|H) = \frac{1}{Z}e^{-\beta H(x)}$

Hamiltonian

Bayes' theorem

 $p(H|Y) \propto p(H)p(Y|H)$

we would like to know

For a set of *iid* cryo-EM particles (not an averaged observable)

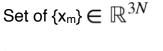
$$p(H|Y) \propto p(H) \prod_{i} \int p(y_i|x_i) p(x_i|H) dx_i$$

• • Approximation: we parametrize the density by a set of $\{x_m\}$ with weights $\{\alpha_m\}$.









Att the the the



 $p(\{\alpha_m\}|Y) \propto p(\{\alpha_m\}) \prod \sum p(y_i|x_m) \alpha_m$ im

Goal: extract the weights



CryoEM images Y={y_i} Approximate probability density

 $p(x|\{\alpha_m\}) = \sum_m \delta(x - x_m)\alpha_m$

Sum to one

*Same expression as in cryoBIFE (Sci Rep 2021), so we can use STAN!

• • Test system Chignolin: Synthetic data

Sufficiently small to have a converged ensemble with MD.

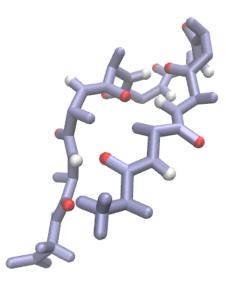
But complex enough with three metastable states:





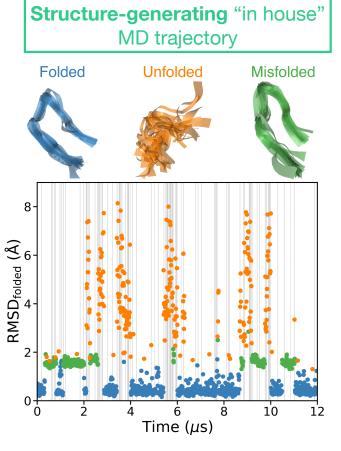






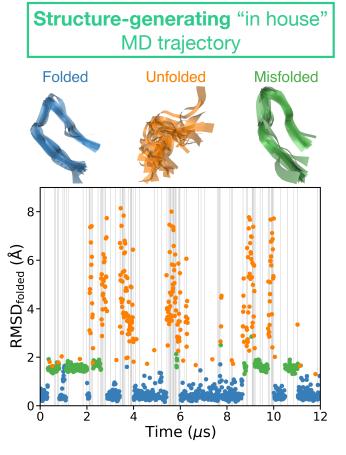


• • Two independent ensembles:



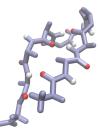
Cluster conformations with K-medoids RMSD (50 centers) = $\{x_m\}$

[○] ○ Two independent ensembles:

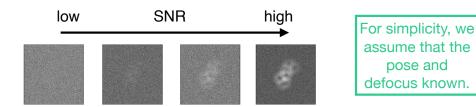


Cluster conformations with K-medoids RMSD (50 centers) = $\{x_m\}$ Image-generating DE Shaw MD trajectory

- -> Different force field
- -> Different ensemble
- -> Structures to generate images are not in the ensemble for refinement.



Example images with different SNR (random pose & defocus)



Can we recover the state populations?

The population is defined as the sum of the weights of the members (i.e., cluster centers) of each state.

		J	
	%fld	%msfd	%unfd
Ground Truth	0.7707	0.0004	0.2289
No noise	0.7784	0.0058	0.2152
SNR = 1.0	0.7786	0.0055	0.2160
SNR = 0.1	0.7787	0.0058	0.2155
SNR = 0.01	0.7693	0.0129	0.2178

Population recovery*:

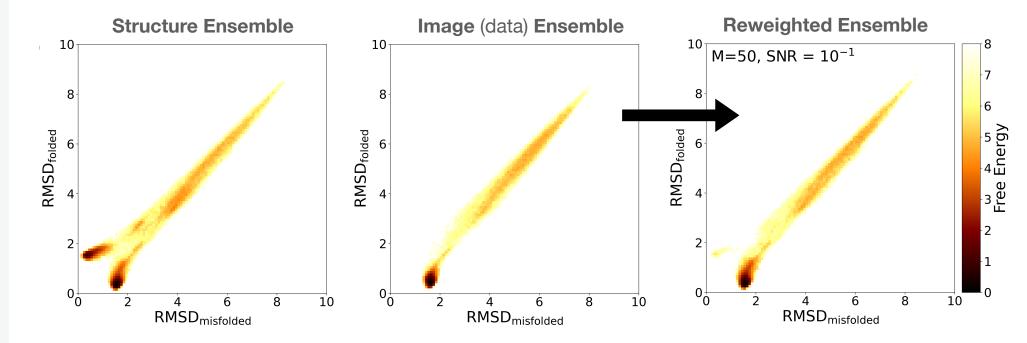
Yes, even at low SNRs*

*If we have to optimize the pose then it becomes more challenging

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Post-processing of the refined ensemble: Free-energy over CVs of choice*.



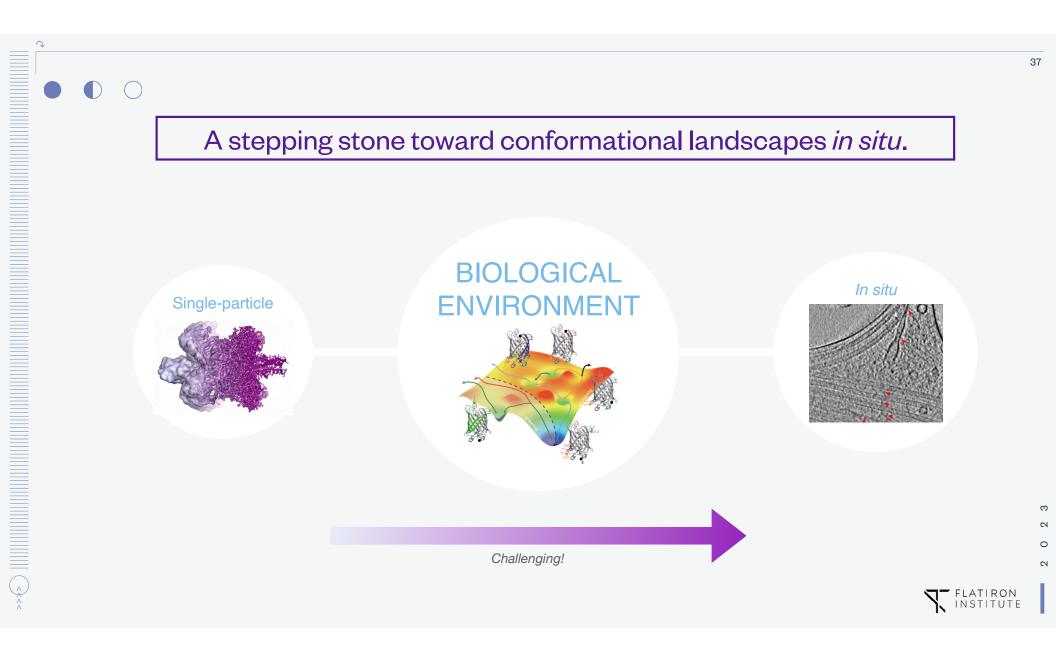
Open questions:

Validation system for conformational variability?

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 How to generate a 'good' ensemble —> bias MD



Acknowledgments

Structural and Molecular Biophysics, CCB/CCM









 \bigcirc \bigcirc

Leslie Greengard



Erik Thiede Gabrie

Alex Barnett

Bob Carpenter Collaborators: Steve Bonilla, Roberto Covino, Attila Szabo, Fabio Pietrucci, Karen Palacio-Rodriguez, Hadrien 2 Vroylandt,.... 0 2

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THANKS FOR YOUR ATTENTION

